Cannabinoid exposure in pubertal rats increases spontaneous ethanol consumption and NMDA receptor associated protein levels

Matthias Klugmann^{1,2}, Viktoria Klippenstein⁴, F. Markus Leweke³, Rainer Spanagel⁴ and Miriam Schneider⁴

¹ Translational Neuroscience Facility, Department of Physiology, School of Medical Sciences, University of New South Wales, Sydney, NSW, Australia

² Department of Physiological Chemistry, University Medical Centre, Johannes Gutenberg University Mainz, Germany

⁸ Department of Psychiatry and Psychotherapy, Central Institute of Mental Health, University of Heidelberg, Germany

⁴ Department of Psychopharmacology, Central Institute of Mental Health, University of Heidelberg, Germany

Abstract

Recent evidence suggests an involvement of the endocannabinoid system in the regulation of emotional behaviour and ethanol intake. Here we investigated age-specific acute behavioural effects of the cannabinoid receptor agonist WIN 55,212-2 (WIN) on anxiety-related behaviour and voluntary ethanol consumption in rats. Animals were treated with WIN (1.2 mg/kg)/vehicle at puberty onset on postnatal day (PD) 40, or at adulthood (PD 100). Animals were tested in the elevated plus-maze (EPM) and the light/ dark emergence test (EMT) and for the initial response to alcohol in a free-choice ethanol consumption paradigm. Acute WIN treatment increased anxiety-related behaviours, and this effect was found to be partially more pronounced in pubertal than adult rats. Additionally, increased intake of higher ethanol solutions after cannabinoid treatment was only observed in pubertal rats. These drug-induced behavioural changes during puberty are paralleled by induction of the NR1 subunit of the NMDA receptor in the medial prefrontal cortex and the striatum. Moreover, pubertal but not adult WIN administration increased the levels of the scaffold protein Homer in these brain regions. Enhanced CB₁ receptor levels in the reinforcement system were also observed in pubertal compared to adult rats. These data support the notion that puberty is a highly vulnerable period for the aversive effects of cannabinoid exposure. In particular, augmented ethanol intake in pubertal cannabinoid-exposed animals might be related to some extent to increased emotional behaviour and in particular to enhanced NMDA and CB1 receptor signalling.

Received 14 June 2010; Reviewed 22 July 2010; Revised 21 October 2010; Accepted 30 November 2010; First published online 7 January 2011

Key words: Anxiety, cannabinoid, ethanol, NMDA, puberty.

Introduction

Products of the hemp plant *Cannabis sativa*, are mainly used for their euphoric and relaxing effects. However, cannabis can also produce dysphoric reactions and states of anxiety (Hall & Solowij, 1998; Patton *et al.* 2002). This apparent paradox is also seen in several animal models where cannabinoids can induce both anxiogenic- and anxiolytic-like responses, depending upon dosage, behavioural paradigms, context

Address for correspondence : Dr M. Schneider, Central Institute of Mental Health, Department of Psychopharmacology,

J5, 68159 Mannheim, Germany.

Tel.: +49 (621) 17036269 Fax: +49 (621) 17036255

(familiar/novel), species and the genetic strain (Valverde, 2005; Viveros *et al.* 2005*b*). However, only few studies have so far investigated if age or developmental status might also influence cannabinoid effects on anxiety-related behaviours.

It has been shown that age of onset of cannabis consumption is an important factor for acute and later consequences of cannabinoid exposure. Consumption/administration of cannabinoids during pubertal development induces lasting cognitive, motivational, social and morphological alterations in humans and rats which are not seen after an identical cannabis use/ treatment in adulthood (Schneider, 2008). Although most of the literature about acute cannabinoid effects in laboratory animals comes from adult rodents, there

ARTICLE

THEMATIC SECTION Consequences of Developmental Exposure to Drugs, Hormones or Altered Environment



Email: miriam.schneider@zi-mannheim.de

are also some results obtained from juvenile and pubertal rats, showing both anxiogenic (McGregor *et al.* 1996; Romero *et al.* 2002; Schneider *et al.* 2008; Schramm-Sapyta *et al.* 2007) and anxiolytic (McGregor *et al.* 1996) effects of acute cannabinoid treatment. Hence, there is only limited data on cannabinoid effects in the context of emotional behaviour during development (Viveros *et al.* 2005*a*).

Recent evidence suggests that the endocannabinoid (ECB) system plays a key role in determining the reinforcing effects of alcohol, since both drugs activate the reward system (for review see Hungund & Basavarajappa, 2004; Mechoulam & Parker, 2003; Spanagel, 2009). The CB₁ receptor antagonist rimonabant (SR141716) has been shown to reduce ethanol intake in rats (Arnone et al. 1997; Colombo et al. 1998; Gallate & McGregor, 1999; Gallate et al. 2004) and cannabinoid agonists increase the motivation for ethanol consumption (Gallate & McGregor, 1999). Furthermore, in CB1 knockout mice reduced ethanol selfadministration (Naassila et al. 2004; Thanos et al. 2005) and ethanol-induced place preference (Houchi et al. 2005; Thanos et al. 2005) has been demonstrated. However, little is known about age-specific actions of cannabinoids on ethanol reward and ethanol intake. So far it has only been shown that an age-dependent decline in ethanol preference observed in aged mice compared to younger mice may be linked to a parallel decline in CB1 receptor signalling (Wang et al. 2003). In general, puberty has been identified as a period associated with higher rates of substance use and abuse (Patton et al. 2004), and since we observed in our previous work stronger acute behavioural cannabinoid effects in pubertal compared to adult rats (Schneider et al. 2008), it might be possible that the modulatory action of the ECB system on ethanol intake is altered as well during pubertal development.

In the present study we therefore investigated the effects of acute administration of the synthetic cannabinoid receptor agonist WIN 55,212-2 (WIN) on anxiety-related behaviour, using the elevated plus-maze (EPM) and the light/dark emergence test (EMT), and on spontaneous ethanol consumption in a free-choice intake paradigm in pubertal [postnatal day (PD) 40] and adult (PD 100) rats.

To correlate the WIN-induced age-dependent behavioural changes with alterations at the molecular level we employed immunoblots to investigate whether acute CB_1 receptor activation would have an impact on the abundance of *N*-methyl-D-aspartate (NMDA) receptor-associated proteins in the reward system [medial prefrontal cortex (mPFC) and the ventral striatum]. The NMDA receptor complex is one of the primary targets of the action of ethanol (Spanagel, 2009; Vengeliene *et al.* 2008) and the composition of post-synaptic scaffolding complexes (e.g. Homer protein expression) in the mesolimbic neurocircuitry plays a crucial role in ethanol-induced neuroplasticity (Obara *et al.* 2009; Szumlinski *et al.* 2005, 2008). To correlate the age-dependent WIN-induced behavioural changes, we assessed protein levels of different NMDA receptor subunits and Homer in pubertal and adult rats following WIN exposure and also quantified the developmental expression profile of the CB₁ receptor.

Methods

Subjects

Male and female Wistar rats were imported from Harlan-Winkelmann (Germany) and housed together in pairs under standard conditions on a 12-h light/ dark cycle (lights on 07:00 hours). They received free access to tap water and were fed *ad libitum*. After 3 wk male rats were removed from the breeding cages. The litters were culled to eight pups directly after birth (all male if possible). In order to avoid litter effects we attempted to assign equal proportions of rats of each litter to the different treatment groups (Zorilla, 1997). A total of 92 first-generation offspring male Wistar rats from our own breeding colony were used for the study.

After weaning on PD 21, male pups were housed in a different room in groups of six (Macrolon cage type IV) under standard conditions on a 12 h light/dark cycle (lights on 06:00 hours). They received free access to tap water and were fed *ad libitum*.

The experiments were performed in accordance with the NIH ethical guidelines and the European Communities Council Directive 86/609/EEC for the care and use of laboratory animals for experiments, and were approved by the local animal care committee (Cologne, Germany).

Drugs

WIN 55,212-2 (WIN) (Sigma-Aldrich, Germany) was dissolved in 0.1% Tween-80 and diluted in saline (0.9%). The drug was administered intraperitoneally at a dose of 1.2 mg/kg. That specific dose was chosen since we have found previously that this concentration of WIN has an effect on behavioural performance (e.g. memory performance and attentional processing) without affecting basic locomotor activity (Drews *et al.* 2005; Schneider & Koch, 2002).

Injection volumes were 1 ml/kg. The experimenter was not aware of the drug treatment of the animals.

Behavioural testing and biochemical analysis was assessed in pubertal (PD 40) and adult (PD 100) rats. Puberty onset in male rats occurs around PD 40 and sexual maturity is reached around PD 60. However, afterwards the animals are still considered to be young adults (late-adolescence) and therefore rats at PD 100 were taken as adult controls (for detailed review see Schneider, 2008). One group of rats was tested for emotional behaviour, starting 30 min after WIN treatment. Therefore, rats were first tested in the light/dark EMT followed 30 min later by the EPM test. Preliminary experiments in our laboratory proved that the sequence of anxiety tests used in the present study had no influence on behaviour during later testing and additionally the effects of WIN on behaviour did not differ if tested 30 or 60 min after injection. Behavioural performance for EPM and EMT was videotaped (digital handycam, Sony, USA) and evaluated offline by a trained experimenter blind to group assignment.

A second cohort of animals was used for investigating the effects of acute WIN treatment on spontaneous alcohol consumption. Alcohol and water were provided 1 h after the cannabinoid injection. Constant background noise was provided by a radio during the light phase of the light/dark cycle in the holding room and for behavioural testing.

For Western blot analysis a third naive group of age-matched animals (PD 40 and PD 100) was used.

Behavioural testing

Light/dark EMT

The EMT took place in a plastic box $(25 \times 80 \times 33 \text{ cm})$ which consisted of two different compartments $(25 \times 40 \times 33 \text{ cm})$, separated by a dividing wall with a hole in the centre that allowed the animals free access to both sides. The first compartment, with black walls could be closed by a lid and was used as the start box. The second compartment had white walls and was brightly illuminated.

Rats were initially placed in the dark, closed compartment and their behaviour was recorded for 5 min after the start box was opened. Subsequent video analysis scored the latency of rats to emerge from the dark compartment into the light compartment, the emergence frequency, the duration of time spent in the light compartment, the amount of rearings and risk assessment behaviour (only head or forepaws are placed in the open compartment without concomitant movement of the hindlimbs, even if the rat subsequently entered the area). The apparatus was thoroughly cleaned with 70% ethanol between sessions.

EPM

The EPM consisted of a plastic plus-shaped apparatus with two open (50×10) and two enclosed arms $(50 \times 10 \times 40 \text{ cm})$, elevated 70 cm above the floor. All arms extended from a central square $(10 \times 10 \text{ cm})$.

At the beginning of each trial, rats were placed on the central square of the plus maze, facing a closed arm. Each rat was tested for 5 min and the following parameters were analysed from videotapes: number of entries into open or closed arms (an entry was defined if all four paws were placed on that arm), time spent in open and closed arms, head dips (the whole head is lowered beneath the edge of an open arm) and risk assessment (only head or forepaws are placed in an open arm without concomitant movement of the hindlimbs, even if the rat subsequently entered the area). Percentage of open arm entries

 $\frac{\text{open arm entries}}{\text{open} + \text{closed arm entries}} \times 100$

and percentage of time spent in open arms

open arm time open+closed arm time ×100

were calculated as well. The total number of closed arm entries was used as an index of general activity. The apparatus was thoroughly cleaned with 70% ethanol between sessions.

Spontaneous ethanol consumption

For the evaluation of ethanol consumption, the animals were singly housed on testing days. Ethanol intake was assessed over a period of 24 h (from 16:00 to 16:00 hours) during which the rats had free access to four different ethanol concentrations (0.5%, 1%, 5%, 10% ethanol) and water. The animals had no previous experience with ethanol, since we were interested in the behaviour upon the first contact with this drug. The volume of ethanol intake was calculated as grams of ethanol/kg of body weight (g/kg) per animal separately for all four different ethanol concentrations and also as total ethanol intake (g/kg) for all concentrations taken together. Additionally, the mean ethanol and water intake during the 24-h period was assessed and ethanol preference was calculated as percentage dependent upon the total fluid intake

100

 $\frac{1}{\text{total fluid intake (ml).}}$ × water or ethanol intake (ml)

Light/dark EMT	Pubertal treatment		Adult treatment	
	Vehicle	WIN	Vehicle	WIN
Emergence frequency	6.5 ± 0.7	4.5 ± 0.8	4.9 ± 1.1	3.3 ± 1.1
Emergence latency (s)	11.7 ± 3.4	77.3 ± 30.9	10.0 ± 2.1	74.4 ± 42.6
Emergence duration (s)	85.4 ± 10.9	46.4 ± 9.1	113.8 ± 24.9	81.0 ± 24.6
Risk assessment	13.1 ± 1.1	11.0 ± 1.1	8.3 ± 0.8	6.0 ± 1.3
Rearing	11.3 ± 1.6	5.1 ± 1.1	11.3 ± 2.3	4.9 ± 1.6

Table 1. Light/dark emergence test (EMT) performance after acute WIN treatment in pubertal and adult rats. A general cannabinoid treatment effect was observed for emergence latency, time spent in the lit compartment and rearing behaviour for pubertal and adult rats, respectively

Data are expressed as mean ± s.E.M.

Western blot analysis

Brain lysate preparation

One hour following administration of WIN or vehicle to adult or pubertal rats, animals were sacrificed and the brains were taken. The ventral striatum and mPFC were dissected and homogenized in 1 ml lysis buffer (10 mM Tris–HCl, 2 mM EDTA; pH 8.0) containing a cocktail of protease inhibitors (Invitrogen, Germany) on crushed ice with a glass homogenizer. In preparation for immunoblotting, aliquots were mixed with an equal volume of $2 \times$ Laemmli reducing sample buffer and heated to 95 °C for 5 min.

Immunoblotting

Using 12% gels, protein samples $(20 \,\mu g)$ were separated by SDS-PAGE, transferred onto PVDF membranes, and probed with the following antibodies:rat anti-EVH1 (panHomer) serum (Klugmann et al. 2005) at 1:1000 dilution, mouse anti-NR1 (1:1000, Millipore, Germany), rabbit anti-NR2b (1:1000, Millipore, Germany), mouse anti- α -tubulin (1:400000, Sigma-Aldrich, Germany), and rabbit anti- β -actin (1:2000; 13E5, Cell Signalling, Germany). For CB1 detection, brain lysates were separated on NuPAGE® Novex bis-tris mini gels 4-12% (Invitrogen, Germany). After transfer, membranes were incubated with the rabbit anti CB₁ receptor antibody (1:500; Cayman Chemical, Estonia). Bound primary antibodies were detected by the appropriate HRP-conjugated secondary antibodies (Dianova, Germany) followed by a chemiluminescent substrate, and the signals captured on film. Densitometric measurement of immunoblot signals was performed using ImageJ software. Signals were standardized to β -actin (CB₁) or tubulin.

Statistical analysis

Treatment effects of the cannabinoid agonist WIN or vehicle on behavioural performance (EPM, EMT, total ethanol intake and preference) and Western blot results in pubertal and adult rats were evaluated using two-way ANOVA, followed by *post-hoc* Tukey *t* tests for pairwise comparisons. Ethanol intake (g/kg) for the different ethanol concentrations was analysed by a two-way ANOVA for repeated measure followed by *post-hoc* Tukey *t* tests for pairwise comparisons. The descriptive statistics was based on means + s.e.m. and a value of p < 0.05 was considered to represent a significant effect.

Results

Light/dark EMT

Statistical analysis revealed a significant WIN treatment effect for emergence latency ($F_{1,38}$ = 4.96, p < 0.05), time spent in the open compartment ($F_{1,38}$ = 4.36, p < 0.05) and rearing behaviour ($F_{1,38}$ = 14.08, p < 0.05) compared to vehicle-treated controls. No treatment effects were observed for risk assessment ($F_{1,38}$ = 3.23, p > 0.05) and emergence frequency ($F_{1,38}$ = 3.71, p < 0.05) (Table 1) (pubertal, WIN group: n = 13; vehicle: n = 12; adult, WIN group: n = 9; vehicle: n = 8).

EPM

Two-way ANOVA revealed a general significant WIN treatment in the EPM effect for time spent in open arms ($F_{1,38}$ =9.25, p<0.05), time spent in closed arms ($F_{1,38}$ =22.01, p<0.05), percentage open arm time ($F_{1,38}$ =10.27, p<0.05) and percentage open arm entries ($F_{1,38}$ =4.47, p<0.05). A significant interaction effect for

Table 2. Effects of acute WIN treatment on Elevated Plus-Maze performance in pubertal and adult rats. A general treatment effect was observed for time spent in open arms, time spent in closed arms, percentage time spent in open arms and percentage open arm entries. Additionally, an age effect was observed for time spent in open and closed arms and percentage open arm time. Significant interaction effects were detected for open arm entries, head dips and risk assessment, with WIN injection inducing only a significant effect in pubertal treated rats (p < 0.05 for age-dependent WIN effects is indicated by asterisks)

	Pubertal treatment		Adult treatment	
EPM	Vehicle	WIN	Vehicle	WIN
Time spent in open arms (s)	13.4 ± 4.3	2.84 ± 1.1	45.8 ± 9.9	20.3 ± 6.2
Time spent in closed arms (s)	255.3 ± 6.3	280.3 ± 3.4	218.4 ± 11.1	263.1 ± 6.8
Time spent in open arms (%)	5.1 ± 1.6	1.1 ± 0.4	17.4 ± 4.7	7.1 ± 2.4
Open arm entries	2.0 ± 0.5	$0.5 \pm 0.2^{*}$	2.8 ± 0.6	2.1 ± 0.6
Closed arm entries	7.3 ± 1.2	6.1 ± 1.3	8.8 ± 1.4	8.7 ± 1.6
Open arm entries (%)	21.2 ± 4.6	10.5 ± 4.3	24.4 ± 4.0	17.1 ± 3.7
Head dips	6.3 ± 0.9	$2.9\pm0.5^*$	5.5 ± 0.7	4.3 ± 0.5
Risk assessment	11.9 ± 1.1	$8.2 \pm 1.1^{*}$	10.5 ± 0.8	9.9 ± 1.2

Data are expressed as mean ± s.E.M.

treatment and age was detected for open arm entries $(F_{1.38} = 4.21, p < 0.05)$, head dips $(F_{1.38} = 4.07, p < 0.05)$ as well as risk assessment ($F_{1,38}$ =4.14, p<0.05). Post-hoc analysis revealed that only pubertal rats showed a significant decrease for open arm entries (p=0.002), head dips (p < 0.001) and risk assessment (p = 0.002) compared to vehicle-treated pubertal controls, whereas no significant difference was observed in the adult treatment groups (p > 0.05). Finally, an age effect was found for time spent in open arms ($F_{1.38} = 17.70$, p < 0.05), time spent in closed arms ($F_{1.38} = 13.17$, p < 1000.05) and percentage open arm time ($F_{1.38} = 16.60$, p < 0.05). No treatment effects were observed on closed arm entries ($F_{1,38} = 0.26$, p > 0.05) (Table 2) (pubertal, WIN group: n = 13; vehicle: n = 12; adult, WIN group: n = 9; vehicle: n = 8).

Spontaneous ethanol consumption

Acute WIN treatment significantly altered ethanol intake (Fig. 1). Statistical analysis revealed a significant interaction effect of treatment and age for the total ethanol intake (g/kg) ($F_{1,28}$ =4.74, p<0.05) (Fig. 1*a*). *Post-hoc* analysis indicated that WIN administration significantly increased ethanol intake specifically in pubertal rats compared to vehicle-treated pubertal (p<0.001) and adult (p<0.001) controls. No such WIN effect was observed in the adult treatment group (p>0.05). Furthermore, within vehicle-treated animals, pubertal rats consumed more ethanol than adult controls (p=0.003). A significant interaction effect was also observed for percentage ethanol preference ($F_{1,28}$ =7.56, p<0.05) (Fig. 1*b*). *Post-hoc* analysis revealed that ethanol preference was significantly increased only in pubertal WIN-treated rats compared to vehicle-treated pubertal controls (p=0.004) and WIN-treated adult rats (p < 0.001). No treatment effects were observed in the adult treatment group (p > 0.05). Finally, when analysing the intake of the different ethanol concentrations (g/kg), a significant interaction effect was observed in pubertal rats (ANOVA: $F_{3,48}$ =2.83; p < 0.05) (Fig. 1*c*) with the intake of the 5% (p=0.001) and 10% (p=0.035) ethanol solutions being significantly increased in cannabinoid-treated animals compared to controls.

No such effects were found in adult rats (ANOVA: $F_{1,30}=0.13$, p>0.05) (Fig. 1*d*) (pubertal, WIN group: n=9; vehicle: n=9; adult, WIN group: n=7; vehicle: n=7).

Biochemical analysis

Western blot for NMDA receptor subunits (NR1 and NR2b) and Homer

Statistical analysis revealed a significant interaction effect between age and cannabinoid treatment for NR1 in the striatum (ANOVA: $F_{1,16}$ =4.75, p <0.05) and the mPFC (ANOVA: $F_{1,16}$ =12.89, p <0.05) (Fig. 2*a*, *b*). *Post-hoc* analysis showed that acute WIN administration significantly increased NR1 expression compared to vehicle-treated controls only in pubertal rats (striatum: p=0.001; mPFC: p=0.021). Furthermore, pubertal WIN-treated rats had significantly higher NR1 levels in both regions compared to adult WIN-treated animals (striatum: p=0.001; mPFC: p=0.002). In addition, in the mPFC adult WIN treatment



Fig. 1. Acute WIN effects on spontaneous ethanol intake in pubertal and adult rats. (*a*) WIN significantly increased the total ethanol intake (g/kg) in pubertal rats compared to vehicle-treated controls and WIN-treated adult animals. Additionally, vehicle-treated pubertal rats were found to have a higher ethanol intake than adult controls. (*b*) Percentage ethanol preference was also specifically increased in pubertal WIN-treated rats only compared to vehicle-treated pubertal controls and WIN-treated adult animals. (*c*) Furthermore, when analysing the ethanol intake of the different ethanol concentrations provided, a significant increase was observed specifically for 5% and 10% ethanol in pubertal rats. (*d*) No such increase was observed in adult animals. Data are expressed as mean + S.E.M., p < 0.05 detected after *post-hoc* analysis are indicated for treatment effects by asterisks (*) and for age effects by daggers (†).

induced a significant reduction in NR1 expression compared to vehicle-treated adult controls (p=0.023). No significant differences in NR1 expression were detected for both regions between vehicle-treated pubertal (Pub) and adult rats (p>0.05) (WIN-Pub, Veh-Pub, WIN-Adult, Veh-Adult: n=5).

Acute WIN treatment did not affect protein levels of NR2b in the striatum (ANOVA : $F_{1,16} = 0.12$, p > 0.05) but we observed a slight but significant (p = 0.023) reduction of NR2B after WIN treatment in the mPCF of adult animals compared to vehicle-treated adult controls ($F_{1,16} = 4.79$, p < 0.05) (Fig. 2*c*, *d*). Furthermore, we also detected an age effect, with vehicle-treated pubertal rats expressing higher NR2B levels than adult animals (p = 0.039) (WIN-Pub, Veh-Pub, WIN-Adult, Veh-Adult: n = 5).

Similar WIN effects as for NR1 could be detected for levels of the NMDA receptor adaptor protein Homer (Fig. 2*e*,*f*). A significant interaction effect of age and

WIN treatment was found for the striatum (ANOVA: $F_{1,16}$ =4.70, p<0.05) and the mPFC (ANOVA: $F_{1,16}$ =8.16, p<0.05). *Post-hoc* analysis revealed that WIN administration significantly increased Homer protein levels exclusively in pubertal rats (striatum: p=0.009; mPFC: p=0.01). In addition, vehicle-treated pubertal rats differed significantly in Homer expression in the striatum from adult controls (p=0.001). In the mPFC cannabinoid-treated pubertal rats showed higher Homer protein levels compared to cannabinoid-treated adults (p=0.001) (WIN-Pub, Veh-Pub, WIN-Adult, Veh-Adult: n=5).

Western blot for CB₁

Statistical analysis detected a significant age effect of CB₁ receptor protein levels between pubertal and adult rats (Fig. 3), independent of WIN treatment, with pubertal rats showing higher CB₁ receptor expression



Fig. 2. Acute effects of WIN in pubertal and adult rats on protein levels of (*a*, *b*) NR1, (*c*, *d*) NR2B and (*e*, *f*) Homer in the striatum and mPFC, respectively, detected by Western blot analysis. Pubertal WIN treatment significantly increased NR1 and Homer levels in the striatum and mPFC. In adult rats a decrease in NR1 and NR2B was specifically detected for the mPFC. Additionally, lower Homer expression levels were found in the striatum of pubertal vehicle-treated rats compared to adult controls. Data are expressed as mean + s.E.M., p < 0.05 after *post-hoc* analysis for treatment effects is indicated by asterisks (*) and for age effects by daggers (†). \blacksquare , WIN; \Box , vehicle.

in the striatum (ANOVA: $F_{1,16}$ =5.15, p<0.05) and mPFC ($F_{1,16}$ =10.46, p<0.05) (WIN-Pub, Veh-Pub, WIN-Adult, Veh-Adult: n=5).

Discussion

In the present study we showed that acute cannabinoid administration in pubertal rats increases

anxiety-related behaviours and the initial consumption and preference of ethanol. Although emotional behaviour was also enhanced after acute WIN treatment in adult rats, no acute effects on ethanol consumption were detected in those animals. These drug-induced behavioural changes observed in pubertal rats were paralleled by induction of the NR1 subunit of the NMDA receptor in the mPFC and striatum. Moreover,



Fig. 3. No acute treatment effects of WIN on CB₁ receptor protein levels were detected in either pubertal or adult rats in the striatum and mPFC. However, an age effect was observed, indicating that the expression of CB₁ receptor levels was significantly higher in all pubertal rats compared to adult groups for both regions. Data are expressed as mean + s.e.m., p < 0.05 for the general age effect is indicated by daggers (†). \blacksquare , WIN; \Box , vehicle.

pubertal but not adult WIN treatment increased the levels of the scaffold protein Homer in these brain regions. These findings are consistent with earlier studies showing that cannabinoids induce stronger effects during pubertal development compared to adult animals (Schneider & Koch, 2003; Schneider, 2008; Schneider *et al.* 2008). Enhanced NMDA receptor complex mediated signalling as well as augmented CB₁ receptor availability in pubertal *vs.* adult animals may contribute to increased vulnerability for the adverse consequences of cannabinoid exposure during puberty.

Emotional behaviour

Acute WIN treatment increased anxiety-like behaviour in pubertal (PD 40) and adult (PD 100) rats in the EPM and the dark/light EMT. However, the anxiogenic effects of WIN were partially more pronounced in pubertal than adult animals. Increasing evidence implicates the involvement of the ECB system in the regulation of anxiety-related behaviour (Moreira & Lutz, 2008; Valverde, 2005; Viveros et al. 2005b; Witkin et al. 2005). Studies investigating age-dependent cannabinoid effects on emotional behaviour are rare and show discrepant findings. In rat pups at PD 12 the cannabinoid agonist CP 55,940 caused a dose-dependent anxiolytic-like reduction of ultrasonic vocalization that was reversed by the CB₁ receptor antagonist rimonabant (McGregor et al. 1996). However, in the same study and also reported by others in pubertal (PD 40) rats (Romero et al. 2002), high doses of CP 55,940

induced hyperreactivity and the rats emitted audible vocalizations when picked up by the experimenter, which might be interpreted as an aversive or anxiogenic-like response. Additionally, it was shown that administration of Δ^9 -tetrahydrocannabinol (THC) in juvenile (PD 28) and late-pubertal (PD 65) male rats increased emotional responses and elevated stress hormone levels for a prolonged period after THC treatment at both ages (Schramm-Sapyta et al. 2007). Unfortunately, no adult controls or pubertal rats were used in this study, which would have indicated the age specifity of these findings. Taken together, these findings indicate that the effects of cannabinoids on emotional behaviour depend to a great extent upon the age of an animal and the developmental state (juvenile, prepubertal, pubertal or early adult).

In humans several reports have shown that early cannabis exposure is significantly associated with later anxiety and depression (Degenhardt *et al.* 2003; Goldschmidt *et al.* 2004; Gray *et al.* 2005; Leech *et al.* 2006; Patton *et al.* 2002; Poulin *et al.* 2005; Rey *et al.* 2002). These studies therefore lend support to our present findings that cannabinoids induce alterations in emotional behaviour in particular during specific vulnerable phases of neuronal development.

Notably, we also observed some age differences between pubertal and adult rats in the EPM, where pubertal rats spent less time in the open arms compared to adult rats. However, since we did not observe any age effects in the light/dark EMT and on other behavioural variables in the EPM such as frequency and percentage of open arm entries and head dips, this difference in open arm time appears not be related primarily to an altered anxiety state in pubertal rats. It is well known that the stress responsiveness is increased during adolescent development (McCormick *et al.* 2010; Romeo, 2010), therefore, it might be possible that the vehicle injection *per se* acted as a stronger stressor in pubertal rats, which might also contribute to the slightly more pronounced WIN effects in pubertal than adult rats.

Initial ethanol intake

In order to assess the initial reaction to ethanol, the intake of different ethanol solutions was measured in alcohol-naive animals. Acute WIN treatment increased ethanol consumption in pubertal rats, in particular the intake of the higher ethanol concentrations (5% and 10%). No such effects were seen in adult animals. Additionally, acute WIN administration increased percentage ethanol preference and the total ethanol intake specifically in pubertal rats.

It has been suggested that the ECB system plays a major role in determining the reinforcing effects of alcohol (Hungund & Basavarajappa, 2004; Mechoulam & Parker, 2003; Spanagel, 2009). The human CB₁ receptor gene (CNR1) may play a role in the development of alcoholism (Benyamina et al. 2011; Zuo et al. 2007). SR141716 reduces ethanol intake in rats (Arnone et al. 1997; Colombo et al. 1998; Gallate & McGregor, 1999; Gallate et al. 2004) and cannabinoid agonists increase the motivation for ethanol consumption (Gallate & McGregor, 1999). Electrophysiological data revealed that the effects of ethanol in the mesolimbic reward pathway relay upon stimulation of CB1 receptors, suggesting the necessity of the ECB system for full development of an appetitive ethanol response (Perra et al. 2005). Furthermore, CB₁ knockout mice show reduced ethanol self-administration (Naassila et al. 2004; Thanos et al. 2005), reduced ethanol-induced place preference (Houchi et al. 2005; Thanos et al. 2005), and ethanol withdrawal symptoms are completely absent in these animals (Racz et al. 2003). In line with these observations we show in the present study that pubertal rats have higher ethanol intake and preference than adult animals, which is paralleled by higher CB₁ receptor protein levels in pubertal rats compared to adults.

Apart from the reinforcing effects of alcohol, several studies indicate that anxiety-related behaviour might be a modulating factor in initial alcohol consumption. It has been suggested that some individuals may be predisposed to alcohol drinking because of high innate anxiety levels (Novak *et al.* 2003; Schuckit &

Hesselbrock, 1994) and anxiety disorders in adolescents are significant predictors of the subsequent onset and persistence of alcohol use and alcoholism (Zimmermann *et al.* 2003). In animal studies it has been shown that elevated anxiety measures in rats correlate with high alcohol consumption during the initiation of alcohol drinking behaviour (Spanagel *et al.* 1995). However, conflicting findings have been reported from rat lines specifically selected and bred for high alcohol intake (Spanagel, 2009). Altogether animal research and epidemiological studies demonstrate the existence of a complex relationship between anxiety-related behaviour and alcohol consumption.

Besides the ethanol intake stimulating effects of WIN in pubertal rats, differences were also observed between pubertal and adult vehicle-treated controls. Total ethanol intake was significantly increased in pubertal compared to adult animals. This finding is consistent with previous studies showing different rewarding effects of ethanol in pubertal than juvenile or adult rats (Philpot et al. 2003) and higher beer intake and binge-like drinking patterns during puberty (Hargreaves et al. 2009). Additionally, adolescent rats are less sensitive to ethanol's aversive properties than adults, measured by conditioned taste aversion (Vetter-O'Hagen et al. 2009). Our finding of enhanced ethanol intake in non-treated pubertal animals, and also the increased stimulatory effects of WIN on pubertal ethanol intake, might be related to some extent to the higher activity of the ECB system at puberty onset (for review see Schneider, 2008) and the higher CB₁ receptor expression observed in the present study.

Biochemical analysis

CB₁ receptors are abundantly expressed in the mPFC and striatum, brain areas known to be involved in emotional behavioural outputs and rewarding effects of drugs such as marijuana or alcohol (Lutz, 2007). In our present study we detected various differences in these brain regions between pubertal and adult rats regarding expression of basal CB₁ receptor protein levels and also reponsiveness towards the acute effects of WIN for NMDA receptor-associated proteins.

Acute WIN treatment did not affect CB_1 receptor expression in both pubertal and adult rats. However, a general age effect was observed with pubertal rats expressing higher protein levels of CB_1 than adults. These results are consistent with an earlier study showing alterations in CB_1 receptor radioligand binding in rats in the limbic forebrain, ventral mesencephalon, and striatum. Binding increased gradually starting from PD 10, and reached maximum values around PD 40 in male animals (Rodriguez de Fonseca *et al.* 1993). Afterwards CB₁ binding was found to be decreased when measured in adult rats at PD 70. These CB₁ receptor dynamics were most pronounced in the ventral mesencephalon and striatum. Therefore, our present results indicate that the heightened receptor sensitivity at the beginning of puberty at PD 40, might be a function of higher innate CB₁ protein levels compared to adulthood. These findings also support our behavioural data where we observed stronger WIN effects in pubertal than in adult rats.

Additionally, we observed an increase in expression of NR1 in the striatum and mPFC after acute WIN treatment of pubertal but not adult rats, whereas NR2B was only altered region-specifically in adult WINtreated animals. A WIN-induced decrease in NR1 and NR2B was detected in the mPFC of adult rats. It is not clear whether these regulations are caused by reduced receptor degradation or increased de novo synthesis. Functional NMDA receptors are assembled from heteromeric subunits including NR1 and at least one of NR2A-D or NR3 (Nakanishi et al. 1998). NR1 is the channel-forming subunit and the NR2 subunits have been proposed to have modulatory effects on channel activity (Monyer et al. 1992). Since the magnitude of WIN-induced decrease for NR2B in adults was less than for NR1, it appears possible that other NR2 subunits may have been co-regulated, thereby contributing to altered NMDA receptor physiology. However, in this study we focused on NR2B because there is compelling evidence that this subunit may constitute a genetic risk factor for stress-induced alcohol drinking and alcoholism (Sillaber et al. 2002). Composition of NMDA receptors containing NR1/NR2B subunits confers high sensitivity to the inhibitory effects of alcohol, as opposed to NR1/NR2C or NR1/NR2D receptors (Allgaier, 2002) and acute alcohol administration inhibits NMDA receptor function (Roberto et al. 2004). Moreover, different studies suggest a role for NR2B-containing rather than NR2A-containing NMDA receptors in modulating the intoxicating effects of ethanol (Boyce-Rustay & Holmes, 2005, 2006). An important role of NR2B in alcohol-induced behaviour is also indicated by the high relative abundance of this subunit in the cortex and striatum compared to other brain regions (Goebel & Poosch, 1999).

The up-regulation of NR1, the major NMDA receptor subunit, after acute CB_1 receptor activiation in pubertal animals suggests enhanced glutamatergic signalling paralleling the initiation of alcohol

consumption. This result is supported by a report on WIN-induced acute activation of NMDA receptors and concomitant release of Ca2+ from intracellular stores in cultured cerebellar granule cells (Netzeband et al. 1999). Although inhibitory effects of cannabinoid function have been described to depress NMDAelicited Ca2+ signals in vitro (Hampson & Deadwyler, 1998), it seems likely that cannabinoid-mediated enhancing effects on NMDA receptor function and intracellular Ca²⁺ might occur under moderate CB₁ receptor stimulation and/or short-term duration. Another explanation for the different effects of WIN in pubertal vs. adult animals might be differences in the molecular composition of constituents of the postsynaptic signalling complex, known as the postsynaptic density.

Homer proteins are critical for regulating the functional architecture of excitatory synapses and long Homer proteins expressing a coiled-coiled domain (CC-Homers) have been shown to recruit group I mGluRs, inositol-1,4,5-triphosphate receptors (IP3Rs) and NR2B to the post-synaptic density. By multimerization they cross-link mGluRs and NMDA receptors to the endoplasmic reticulum membrane and modulate intracellular Ca²⁺ dynamics (Klugmann & Szumlinski, 2008). This adapter function makes Homer an attractive candidate for enabling a functional interplay between the CB1 and NMDA receptor signalling pathways. Indeed, direct involvement of Homer in ECB-mediated neuroplasticity has been indicated in recent studies. NMDA receptors are connected via PSD-95, GKAP, and SHANK to Homer that in turn binds to proteins involved in ECB signalling such as mGluRs and diacylglycerol lipase (DGL) (Klugmann & Szumlinski, 2008). Jung et al. (2007) have shown that Homer proteins enhance retrograde ECB signalling by promoting plasma membrane expression of DGL- α in a neuroblastoma cell line. Moreover, the cocaineinduced attenuation of retrograde ECB signalling known to underlie long-term depression has been correlated with enhanced expression levels of Homer proteins (Fourgeaud et al. 2004). Interestingly, the induction of ECB-mediated synaptic plasticity is also induced after stimulation of the immediate early gene isoform, Homer 1a. This isoform, known to be induced by neuronal activity (Brakeman et al. 1997), suppresses metabotropic-mediated but enhances depolarizationmediated endocannabinoid production (Roloff et al. 2010).

In the present study, we detected an age-specific induction of NR1 and CC-Homer after acute WIN treatment specifically in pubertal rats. Importantly, CC-Homer proteins in corticolimbic brain regions have been shown to be critical for emotional behaviour (Klugmann & Szumlinski, 2008). Their regulation by various drugs of abuse is not only subregionally selective, but also drug-specific and isoform-specific. In the context of alcohol, Homer2a/b is up-regulated within the accumbens, but Homer1b/c is unchanged (Obara et al. 2009; Szumlinski et al. 2008). In the context of cocaine, both Homer1b/c and Homer2a/b are downregulated within the accumbens, but Homer2a/b is up-regulated in the PFC (Ary & Szumlinski, 2007; Ben Shahar et al. 2009; Swanson et al. 2001). Moreover, these effects are observed 24 h (alcohol studies) or 3 wk (cocaine studies) following the last drug exposure. It is not known whether or not CC-Homers or their associated glutamate receptors change within the brain at shorter withdrawal time-points (i.e. at 1 h postinjection as observed in the present study). NR1 and long Homer isoforms are constitutively expressed, and it is not clear what cellular mechanisms might have accounted for short-term changes in the respective protein levels. It appears intuitive that NR1 levels may be increased by a reduced turnover or internalization while transcriptional or translational regulations can not be excluded. Our data suggest that relatively quick changes in NR1 and CC-Homer protein expression can occur following acute drug treatment that are likely to affect glutamate signalling of relevance to drug reward.

Interestingly, we observed lower CC-Homer protein expression in the striatum of vehicle-treated pubertal than adult rats. This result matches the reported developmental expression profile of Homer 1 and 2 in different brain regions, including the striatum (Shiraishi *et al.* 2004), where increasing protein levels were detected throughout the early postnatal developmental period in mice.

Conclusion

We were able to demonstrate with the present study that administration of a cannabinoid agonist induces stronger behavioural effects in pubertal rats compared to adults, by increasing anxiety-related behaviours and initial ethanol intake. These behavioural alterations were paralleled by differential inductions of NMDA receptor-associated proteins in pubertal and adult rats. The present data further demonstrate the susceptibility of the pubertal developmental period for the aversive behavioural consequences of cannabinoid exposure, which might be related to enhanced activity of the ECB system and concomitant alterations in glutamatergic signalling during this specific developmental period.

Acknowledgements

We thank Theresa Brand and Ruth Jelinek for excellent technical assistance. Miriam Schneider was supported by DFG (SCHN 958/3–1).

Statement of Interest

None.

References

- Allgaier C (2002). Ethanol sensitivity of NMDA receptors. Neurochemistry International 41, 377–382.
- Arnone M, Maruani J, Chaperon F, Thiebot M, *et al.* (1997). Selective inhibition of sucrose and ethanol intake by SR 1411716, an antagonist of central cannabinoid (CB1) receptors. *Psychopharmacology* **132**, 104–106.
- Ary AW, Szumlinski KK (2007). Regional differences in the effects of withdrawal from repeated cocaine upon Homer and glutamate receptor expression: a two-species comparison. *Brain Research* 1184, 295–305.
- Ben Shahar O, Obara I, Ary AW, Ma N, et al. (2009). Extended daily access to cocaine results in distinct alterations in Homer 1b/c and NMDA receptor subunit expression within the medial prefrontal cortex. Synapse 63, 598–609.
- Benyamina A, Kebir O, Blecha L, Reynaud M, *et al.* (2011). CNR1 gene polymorphisms in addictive disorders: a systematic review and a meta-analysis. *Addiction Biology* **16**, 1–6.
- **Boyce-Rustay JM, Holmes A** (2005). Functional roles of NMDA receptor NR2A and NR2B subunits in the acute intoxicating effects of ethanol in mice. *Synapse* **56**, 222–225.
- **Boyce-Rustay JM, Holmes A** (2006). Ethanol-related behaviors in mice lacking the NMDA receptor NR2A subunit. *Psychopharmacology* (*Berlin*) **187**, 455–466.
- Brakeman PR, Lanahan AA, O'Brien R, Roche K, *et al.* (1997). Homer: a protein that selectively binds
- metabotropic glutamate receptors. *Nature* **386**, 284–288. **Colombo G, Agabio R, Fa M, Guano L**, *et al.* (1998). Reduction of voluntary ethanol intake in ethanolpreferring sP rats by the cannabinoid antagonist SR141716.
- Alcohol and Alcoholism 33, 126–130.
 Degenhardt L, Hall W, Lynskey M (2003). Exploring the association between cannabis use and depression.
 Addiction 98, 1493–1504.
- Drews E, Schneider M, Koch M (2005). Effects of the cannabinoid receptor agonist WIN 55,212-2 on operant behavior and locomotor activity in rats. *Pharmacology Biochemistry and Behavior* 80, 145–150.
- Fourgeaud L, Mato S, Bouchet D, Hemar A, et al. (2004). A single *in vivo* exposure to cocaine abolishes endocannabinoid-mediated long-term depression in the nucleus accumbens. *Journal of Neuroscience* 24, 6939–6945.

Gallate JE, Mallet PE, McGregor IS (2004). Combined low dose treatment with opioid and cannabinoid receptor antagonists synergistically reduces the motivation to consume alcohol in rats. *Psychopharmacology* **173**, 210–216.

Gallate JE, McGregor IS (1999). The motivation for beer in rats: effects of ritanserin, naloxone and SR141716. *Psychopharmacology* **142**, 302–308.

Goebel DJ, Poosch MS (1999). NMDA receptor subunit gene expression in the rat brain: a quantitative analysis of endogenous mRNA levels of NR1Com, NR2A, NR2B, NR2C, NR2D and NR3A. *Brain Research* 69, 164–170.

Goldschmidt L, Richardson GA, Cornelius MD, Day NL (2004). Prenatal marijuana and alcohol exposure and academic achievement at age 10. *Neurotoxicology and Teratology* **26**, 521–532.

Gray KA, Day NL, Leech S, Richardson GA (2005). Prenatal marijuana exposure: effect on child depressive symptoms at ten years of age. *Neurotoxicology and Teratology* 27, 439–448.

Hall W, Solowij N (1998). Adverse effects of cannabis. *Lancet* 352, 1611–1616.

Hampson RE, Deadwyler SA (1998). Role of cannabinoid receptors in memory storage. *Neurobiology of Disease* 5, 474–482.

Hargreaves GA, Monds L, Gunasekaran N, Dawson B, et al. (2009). Intermittent access to beer promotes binge-like drinking in adolescent but not adult Wistar rats. *Alcohol* **43**, 305–314.

Houchi H, Babovic D, Pierrefiche O, Ledent C, *et al.* (2005). CB1 receptor knockout mice display reduced ethanolinduced conditioned place preference and increased striatal dopamine D2 receptors. *Neuropsychopharmacology* **30**, 339–349.

Hungund BL, Basavarajappa BS (2004). Role of endocannabinoids and cannabinoid CB1 receptors in alcohol-related behaviors. *Annals of New York Academy of Sciences* **1025**, 515–527.

Jung KM, Astarita G, Zhu C, Wallace M, *et al.* (2007). A key role for diacylglycerol lipase-alpha in metabotropic glutamate receptor-dependent endocannabinoid mobilization. *Molecular Pharmacology* **72**, 612–621.

Klugmann M, Symes CW, Leichtlein CB, Klaussner BK, *et al.* (2005). AAV-mediated hippocampal expression of short and long Homer 1 proteins differentially affect cognition and seizure activity in adult rats. *Molecular and Cellular Neuroscience* **28**, 347–360.

Klugmann M, Szumlinski KK (2008). Targeting Homer genes using adeno-associated viral vector: lessons learned from behavioural and neurochemical studies. *Behavioural Pharmacology* **19**, 485–500.

Leech SL, Larkby CA, Day R, Day NL (2006). Predictors and correlates of high levels of depression and anxiety symptoms among children at age 10. *Journal of the American Academy of Child Adolescent Psychiatry* **45**, 223–230.

Lutz B (2007). The endocannabinoid system and extinction learning. *Molecular Neurobiology* **36**, 92–101.

McCormick CM, Mathews IZ, Thomas C, Waters P (2010). Investigations of HPA function and the enduring consequences of stressors in adolescence in animal models. *Brain and Cognition* **72**, 73–85.

McGregor IS, Dastur FN, McLellan RA, Brown RE (1996). Cannabinoid modulation of rat pup ultrasonic vocalizations. *European Journal of Pharmacology* **313**, 43–49.

Mechoulam R, Parker L (2003). Cannabis and alcohol – a close friendship. *Trends in Pharmacological Sciences* 24, 266–268.

Monyer H, Sprengel R, Schoepfer R, Herb A, et al. (1992). Heteromeric NMDA receptors: molecular and functional distinction of subtypes. *Science* **256**, 1217–1221.

Moreira FA, Lutz B (2008). The endocannabinoid system: emotion, learning and addiction. *Addiction Biology* **13**, 196–212.

Naassila M, Pierrefiche O, Ledent C, Daoust M (2004). Decreased alcohol self-administration and increased alcohol sensitivity and withdrawal in CB1 receptor knockout mice. *Neuropharmacology* **46**, 243–253.

Nakanishi S, Nakajima Y, Masu M, Ueda Y, *et al.* (1998). Glutamate receptors: brain function and signal transduction. *Brain Research Reviews* **26**, 230–235.

Netzeband JG, Conroy SM, Parsons KL, Gruol DL (1999). Cannabinoids enhance NMDA-elicited Ca2 + signals in cerebellar granule neurons in culture. *Journal of Neuroscience* **19**, 8765–8777.

Novak A, Burgess ES, Clark M, Zvolensky MJ, et al. (2003). Anxiety sensitivity, self-reported motives for alcohol and nicotine use, and level of consumption. *Journal of Anxiety Disorders* 17, 165–180.

Obara I, Bell RL, Goulding SP, Reyes CM, *et al.* (2009). Differential effects of chronic ethanol consumption and withdrawal on homer/glutamate receptor expression in subregions of the accumbens and amygdala of P rats. *Alcoholism: Clinical Experimental Research* **33**, 1924–1934.

Patton GC, Coffey C, Carlin JB, Degenhardt L, et al. (2002). Cannabis use and mental health in young people: cohort study. British Medical Journal 325, 1195–1198.

Patton GC, McMorris BJ, Toumbourou JW, Hemphill SA, *et al.* (2004). Puberty and the onset of substance use and abuse. *Pediatrics* **114**, e300–e306.

Perra S, Pillolla G, Melis M, Muntoni AL, et al. (2005). Involvement of the endogenous cannabinoid system in the effects of alcohol in the mesolimbic reward circuit: electrophysiological evidence *in vivo*. *Psychopharmacology* 183, 368–377.

Philpot RM, Badanich KA, Kirstein CL (2003). Place conditioning: age-related changes in the rewarding and aversive effects of alcohol. *Alcoholism: Clinical Experimental Research* 27, 593–599.

Poulin C, Hand D, Boudreau B, Santor D (2005). Gender differences in the association between substance use and elevated depressive symptoms in a general adolescent population. *Addiction* 100, 525–535.

 Racz I, Bilkei-Gorzo A, Toth ZE, Michel K, et al. (2003).
 A critical role for the cannabinoid CB1 receptors in alcohol dependence and stress-stimulated ethanol drinking. Journal of Neuroscience 23, 2453–2458. Rey JM, Sawyer MG, Raphael B, Patton GC, et al. (2002). Mental health of teenagers who use cannabis. Results of an Australian survey. *British Journal of Psychiatry* **180**, 216–221.

Roberto M, Schweitzer P, Madamba SG, Stouffer DG, et al. (2004). Acute and chronic ethanol alter glutamatergic transmission in rat central amygdala: an *in vitro* and *in vivo* analysis. *Journal of Neuroscience* 24, 1594–1603.

Rodriguez de Fonseca F, Ramos JA, Bonnin A, Fernandez-Ruiz JJ (1993). Presence of cannabinoid binding sites in the brain from early postnatal ages. *Neuroreport* 4, 135–138.

Roloff AM, Anderson GR, Martemyanov KA, Thayer SA (2010). Homer 1a gates the induction mechanism for endocannabinoid-mediated synaptic plasticity. *Journal of Neuroscience* **30**, 3072–3081.

Romeo RD (2010). Adolescence: a central event in shaping stress reactivity. *Developmental Psychobiology* **52**, 244–253.

Romero EM, Fernandez B, Sagredo O, Gomez N, *et al.* (2002). Antinociceptive, behavioural and neuroendocrine effects of CP 55,940 in young rats. *Brain Research, Developmental Brain Research* **136**, 85–92.

Schneider M (2008). Puberty as a highly vulnerable developmental period for the consequences of cannabis exposure. *Addiction Biology* **13**, 253–263.

Schneider M, Koch M (2002). The cannabinoid agonist WIN 55,212-2 reduces sensorimotor gating and recognition memory in rats. *Behavioral Pharmacology* 13, 29–37.

Schneider M, Koch M (2003). Chronic pubertal, but not adult chronic cannabinoid treatment impairs sensorimotor gating, recognition memory and the performance in a progressive ratio task in adult rats. *Neuropsychopharmacology* **28**, 1760–1769.

Schneider M, Schomig E, Leweke FM (2008). Acute and chronic cannabinoid treatment differentially affects recognition memory and social behavior in pubertal and adult rats. *Addiction Biology* **13**, 345–357.

Schramm-Sapyta NL, Cha YM, Chaudhry S, Wilson WA, et al. (2007). Differential anxiogenic, aversive, and locomotor effects of THC in adolescent and adult rats. *Psychopharmacology* **191**, 867–877.

Schuckit MA, Hesselbrock V (1994). Alcohol dependence and anxiety disorders: what is the relationship? *American Journal of Psychiatry* **151**, 1723–1734.

Shiraishi Y, Mizutani A, Yuasa S, Mikoshiba K, et al. (2004). Differential expression of Homer family proteins in the developing mouse brain. *Journal of Comparative Neurology* 473, 582–599.

Sillaber I, Rammes G, Zimmermann S, Mahal B, et al. (2002). Enhanced and delayed stress-induced alcohol drinking in mice lacking functional CRH1 receptors. *Science* **296**, 931–933.

Spanagel R (2009). Alcoholism: a systems approach from molecular physiology to addictive behavior. *Physiological Reviews* 89, 649–705.

Spanagel R, Montkowski A, Allingham K, Stohr T, *et al.* (1995). Anxiety: a potential predictor of vulnerability to the

initiation of ethanol self-administration in rats. *Psychopharmacology* (*Berlin*) **122**, 369–373.

Swanson CJ, Baker DA, Carson D, Worley PF, et al. (2001). Repeated cocaine administration attenuates group I metabotropic glutamate receptor-mediated glutamate release and behavioral activation: a potential role for Homer. *Journal of Neuroscience* **21**, 9043–9052.

Szumlinski KK, Ary AW, Lominac KD, Klugmann M, et al. (2008). Accumbens Homer2 overexpression facilitates alcohol-induced neuroplasticity in C57BL/6J mice. *Neuropsychopharmacology* **33**, 1365–1378.

Szumlinski KK, Lominac KD, Oleson EB, Walker JK, et al. (2005). Homer2 is necessary for EtOH-induced neuroplasticity. Journal of Neuroscience 25, 7054–7061.

Thanos PK, Dimitrakakis ES, Rice O, Gifford A, et al. (2005). Ethanol self-administration and ethanol conditioned place preference are reduced in mice lacking cannabinoid CB1 receptors. *Behavioural Brain Research* 164, 206–213.

Valverde O (2005). Participation of the cannabinoid system in the regulation of emotional-like behaviour. *Current Pharmaceutical Design* **11**, 3421–3429.

Vengeliene V, Bilbao A, Molander A, Spanagel R (2008). Neuropharmacology of alcohol addiction. *British Journal of Pharmacology* 154, 299–315.

Vetter-O'Hagen C, Varlinskaya E, Spear L (2009). Sex differences in ethanol intake and sensitivity to aversive effects during adolescence and adulthood. *Alcohol and Alcoholism* 44, 547–554.

Viveros MP, Llorente R, Moreno E, Marco EM (2005*a*). Behavioural and neuroendocrine effects of cannabinoids in critical developmental periods. *Behavioral Pharmacology* 16, 353–362.

Viveros MP, Marco EM, File SE (2005b). Endocannabinoid system and stress and anxiety responses. *Pharmacology Biochemistry Behavior* 81, 331–342.

Wang L, Liu J, Harvey-White J, Zimmer A, et al. (2003). Endocannabinoid signaling via cannabinoid receptor 1 is involved in ethanol preference and its age-dependent decline in mice. Proceedings of the National Academy of Sciences USA 100, 1393–1398.

Witkin JM, Tzavara ET, Nomikos GG (2005). A role for cannabinoid CB1 receptors in mood and anxiety disorders. *Behavioral Pharmacology* **16**, 315–331.

Zimmermann P, Wittchen HU, Hofler M, Pfister H, *et al.* (2003). Primary anxiety disorders and the development of subsequent alcohol use disorders: a 4-year community study of adolescents and young adults. *Psychological Medicine* **33**, 1211–1222.

Zorilla EP (1997). Multiparous species present problems (and possibilities) to developmentalists. *Developmental Psychobiology* **30**, 141–150.

Zuo L, Kranzler HR, Luo X, Covault J, *et al.* (2007). CNR1 variation modulates risk for drug and alcohol dependence. *Biological Psychiatry* **62**, 616–626.